

This article was downloaded by:

On: 18 January 2011

Access details: Access Details: Free Access

Publisher Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



International Journal of Environmental Analytical Chemistry

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713640455>

The Nonreversibility of Adsorption-Desorption of Dieldrin and Lindane on Chitin in Seawater

J. M. Santana-casiano^a; J. Perez-Peña^a; M. Gonzalez-davila^a

^a Departamento de Química, Universidad de Las Palmas de Gran Canaria, Facultad de Ciencias del Mar., Las Palmas, España

To cite this Article Santana-casiano, J. M. , Perez-Peña, J. and Gonzalez-davila, M.(1992) 'The Nonreversibility of Adsorption-Desorption of Dieldrin and Lindane on Chitin in Seawater', International Journal of Environmental Analytical Chemistry, 46: 1, 163 — 174

To link to this Article: DOI: 10.1080/03067319208027007

URL: <http://dx.doi.org/10.1080/03067319208027007>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

THE NONREVERSIBILITY OF ADSORPTION-DESORPTION OF DIELDRIN AND LINDANE ON CHITIN IN SEAWATER

J. M. SANTANA-CASIANO, J. PEREZ-PEÑA and M. GONZALEZ-DAVILA

*Universidad de Las Palmas de Gran Canaria. Facultad de Ciencias del Mar.
Departamento de Química. P.O. Box 550. Las Palmas. España.*

(Received 13 March 1991; in final form, 31 July 1991)

Adsorption and desorption of dieldrin and lindane on chitin were investigated in seawater batch tests as a function of chitin concentration, temperature, pH and salinity. For chitin concentrations ranging from 0.5 g l^{-1} to 12.5 g l^{-1} , the pesticide concentrations were varied from $4 \mu\text{g l}^{-1}$ to $65 \mu\text{g l}^{-1}$ for dieldrin and from $40 \mu\text{g l}^{-1}$ to $680 \mu\text{g l}^{-1}$ for lindane. Both dieldrin and lindane show adsorption-desorption hysteresis at low chitin concentration. At high chitin concentrations ($m > 6.25 \text{ g l}^{-1}$ for dieldrin and $m > 10 \text{ g l}^{-1}$ for lindane) both pesticides exhibit reversible behaviour. However, only lindane adsorption is affected by chitin concentration. These types of behaviour remain fixed in prewashed chitin. However, an increase in the temperature and a decrease in the salinity made the process become reversible. A resistant-reversible two component model has been applied to account for these types of behaviour and provides a way to explain most of the observed effects by defining mass independent distribution coefficients.

KEY WORDS: Adsorption, desorption, dieldrin, lindane, chitin, seawater.

INTRODUCTION

In aquatic environments, one of the fundamental processes controlling the behaviour, fate and distribution of persistent organic compounds is the exchange of these chemicals between dissolved and sorbed states. Clay and suspended organic particulate matter are considered important for adsorption, but very little is known about the actual adsorption-desorption process¹⁻³. The scavenging of chlorinated hydrocarbons by organic particulates could provide a means of transporting these persistent pollutants through the water column and for concentrating them in sediments and in benthic and planktonic detritus-feeding organisms. The strong particle-binding tendency of persistent organic chemicals such as DDT, lindane, and PCBs is well known, and its importance has been stressed many times^{4,5}.

Chitin (poly-*N*-acetyl-D-glucosamine), as well as its deacetylated derivative chitosan, is a constituent of the natural dissolved and particulate organics and part of fungi and bacteria. It is consequently abundant in the natural environment. The extent of chlorinated hydrocarbon sorption on chitin is, thus, a critical component in the evaluation of its fate in water systems. However, very little is known about the interaction of pesticides with this polymer^{6,7}.

In this work we present the results of the adsorption-desorption of dieldrin and lindane on chitin in seawater. The studies have been done taking into account two important facts of these processes: first, the reversibility or nonreversibility of the adsorption reaction^{2,8} between the lindane and dieldrin and the natural chitin, and second, the effect of chitin concentration on the adsorption-desorption process^{3,9,10}. Results of studies at different pH, temperature, and salinity values were also analysed.

EXPERIMENTAL

The chitin powder used was derived from crab shells and obtained from Sigma (Sigma, Chemical Co., MO, U.S.A.). The surface area ($0.5 \pm 0.03 \text{ m}^2 \text{ g}^{-1}$) for the bulk chitin and the amphoteric properties of the surface of chitin ($\text{pK}_{a1} = 4.4 \pm 0.2$ and $\text{pK}_{a2} = 6.4 \pm 0.3$) were determined in a previous work¹¹. Preliminary adsorption studies with various size fractions from less than $88 \mu\text{m}$ to $250 \mu\text{m}$ give similar values to those of the bulk, which was used in all the following studies. Stock solutions of dieldrin (98%) and lindane (99%) (Supelco Inc, Bellefonte, PA, U.S.A.) were prepared in pesticide grade hexane 7.5 ppm and 75 ppm, respectively.

All batch sorption studies (in triplicate) were conducted by preequilibrating the dieldrin (ranging from $4 \mu\text{g l}^{-1}$ to $65 \mu\text{g l}^{-1}$) (solubility¹² $2 \cdot 10^2 \mu\text{g l}^{-1}$) and the lindane (ranging from $40 \mu\text{g l}^{-1}$ to $685 \mu\text{g l}^{-1}$) (solubility¹² $7.8 \cdot 10^3 \mu\text{g l}^{-1}$) with 40 ml of $0.45 \mu\text{m}$ filtered and hexane-preeextracted seawater ($S = 36.52\%$) from Gran Canaria (Canary Islands). After 24 h preequilibration period, the chitin was added (from 0.5 g l^{-1} to 12.5 g l^{-1}) and the glass stoppered centrifuge glass tubes (volume: 50 cm^3) were shaken in a controlled temperature bath for 24 h adsorption and 24 h desorption time periods when equilibrium conditions were desired. After shaking, the supernatant was centrifuged for 15 min at 6,000 rpm, after which 25 ml of supernatant was withdrawn and extracted with 10 ml of hexane. In the recovery studies, repeated extractions showed the first extraction removed over 97% of both pesticide masses. In all the dieldrin studies, the extract was reduced to 1 ml in rotavapor at 40°C and at 360 mm Hg controlled vacuum pressure. Extracted samples were analysed and quantified by gas chromatography with a linearized ^{63}Ni ECD detector. A SPB-5 (Supelco Inc) 30 m-fused silica capillary column was used with N_2 carrier gas at a linear velocity of 25 cm s^{-1} . Samples were analysed isothermally (200°C) using on-column injection and a detector temperature of 300°C .

The pesticide mass was derived from triplicate measurements by comparing their peak area responses to a standard curve prepared with standard solution of the pesticide in the same experimental conditions. The coefficients of variation of these triplicate measurements were about 1%. It was assumed that the difference between the pesticide mass added and the pesticide mass in liquid phase at the end was the result of chitin adsorption.

For the desorption, the following procedure was repeated several times: the supernatant was replaced with contaminant-free seawater, shaken again for 24 hour periods, when equilibrium conditions were desired, recentrifuged, and the new supernatant analysed.

RESULTS AND DISCUSSION

Adsorption-desorption kinetics

The adsorption-desorption rate of dieldrin and lindane on chitin (2 g l^{-1} and 6.25 g l^{-1} , respectively) was determined at initial concentrations of $18.75 \mu\text{g l}^{-1}$ (dieldrin) and $187.5 \mu\text{g l}^{-1}$ (lindane) in seawater. For the adsorption studies, pesticide solutions were equilibrated with the chitin for periods ranging from 5 min to 92 h and for the desorption studies, after 24 h equilibration periods, the contaminated chitin was preequilibrated with contaminant-free seawater for periods ranging from 5 to 74 hr. A comparison of the lindane and the dieldrin adsorbed and desorbed from solution by chitin is shown in Figure 1. The data show that initial uptake is rapid mainly for lindane and a 3 h period for the adsorption of lindane and 10 h periods for the adsorption of dieldrin were enough to reach equilibrium. For the desorption experiments, the process is essentially complete in a few minutes and much longer desorption times do not produce notable effects. 24 h equilibrium periods both for adsorption and desorption experiments were used in all the studies. The present results do suggest kinetically rapid adsorption-desorption reactions, similar to other studies on organic partitioning between the sediment and water phases^{2,13}.

Isotherms for dieldrin and lindane

Figure 2 shows the results of single adsorption-desorption of dieldrin conducted at two chitin concentration and this is typical of all the results obtained over the study.

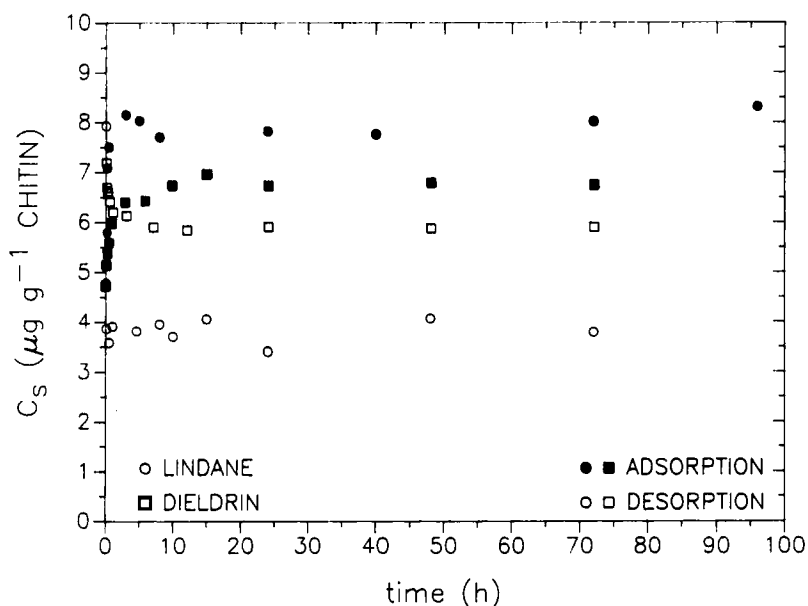


Figure 1 Adsorption-desorption kinetics of dieldrin and lindane on chitin. For the desorption kinetics, adsorption equilibrium time was fixed at 24 h.

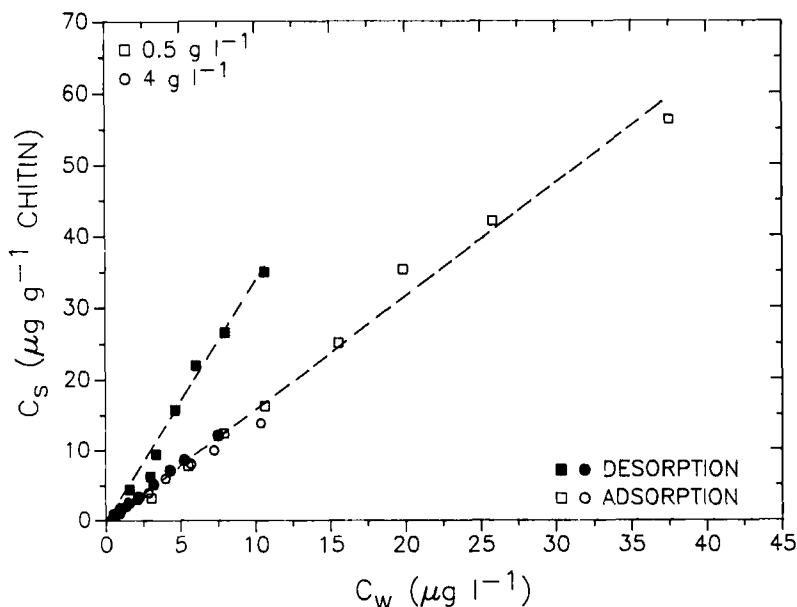


Figure 2 Dieldrin adsorption-desorption isotherms on chitin ($m = 0.5 \text{ g l}^{-1}$ and $m = 4 \text{ g l}^{-1}$). Data are averages of three replicates.

The behaviour shown by the lindane adsorption-desorption is also depicted in Figure 3. Around 60% of dieldrin is adsorbed per gram of chitin and it is 25-times more strongly adsorbed on chitin than lindane. Only 25% on average of the lindane dissolved in seawater is adsorbed per gram of chitin. The increased uptake of dieldrin over lindane may be due to the asymmetry of the dieldrin molecule which may influence its electron density causing increased polarization compared to lindane. The increased polarity may result in stronger binding forces between the dieldrin and the chitin particles.

Reasonable linearity over the range of concentrations studied can be observed for both pesticides, and the near zero intercept of a linear regression of the data supports the validity of a linear partitioning model.

$$C_{sa} = K_a C_{wa} \quad (1)$$

where C_{sa} and C_{wa} are the adsorbed and aqueous equilibrium concentration and K_a is the partition coefficient for the adsorption.

Similarly, the desorption points fall on a straight line, and a desorption isotherm which corresponds with the previous adsorption reaction, can be defined as

$$C_{sd} = K_d C_{wd} \quad (2)$$

with partition coefficient, K_d , and C_{sd} and C_{wd} the adsorbed and aqueous equilibrium concentration for the desorption studies. The results illustrated in Figures 2 and 3

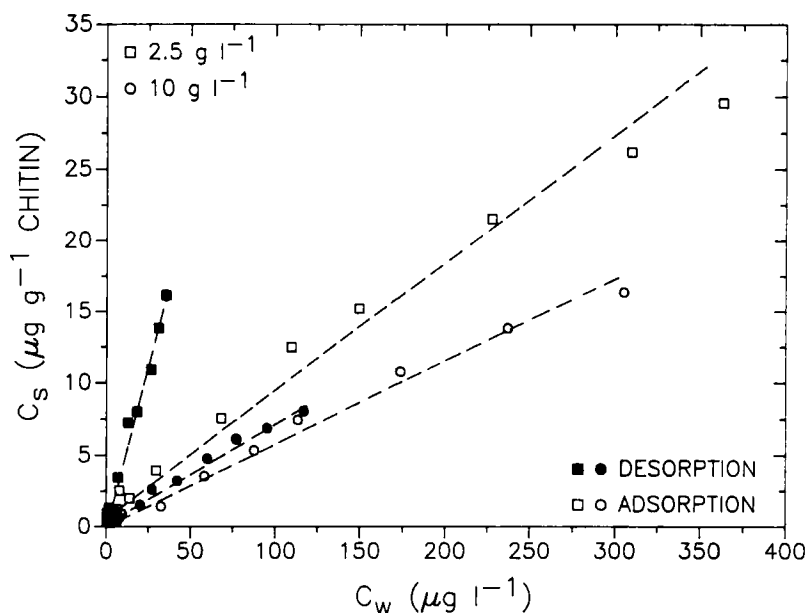


Figure 3 Adsorption-desorption isotherms of lindane at two chitin concentrations ($m = 2.5 \text{ g l}^{-1}$ and $m = 10 \text{ g l}^{-1}$). The sorption is not reversible and there exists an adsorbent mass effect.

for dieldrin and lindane adsorption-desorption at the different chitin concentrations studied in seawater are summarized in Table 1 and 2, respectively.

Two important facts can be extracted from Figures 2 and 3 and Tables 1 and 2. First, adsorption and subsequent desorption experiments for both dieldrin and lindane appear to demonstrate only partially reversible behaviour. That is, the measured desorption partition coefficients do not follow the adsorption partition

Table 1 Adsorption and desorption partition coefficients and the resistant-reversible component partition coefficients for the adsorption of dieldrin to chitin in seawater (temperature 22°C , salinity 36.52‰)

$[Chitin](\text{g l}^{-1})$	$K_a(\text{l g}^{-1})$	$K_d(\text{l g}^{-1})$	$K_o(\text{l g}^{-1})$	$K_x(\text{l g}^{-1})$
0.5	1.58 ± 0.07	3.55 ± 0.17	0.74 ± 0.03	0.72 ± 0.02
1	1.58 ± 0.05	2.14 ± 0.04	0.48 ± 0.03	1.02 ± 0.03
1* ¹	1.82 ± 0.08	3.01 ± 0.08	1.09 ± 0.02	0.82 ± 0.04
1* ²	1.05 ± 0.04	1.34 ± 0.06	0.29 ± 0.03	0.96 ± 0.04
1* ¹	1.56 ± 0.06	1.90 ± 0.05	0.19 ± 0.03	1.30 ± 0.04
1* ²	1.60 ± 0.05	1.60 ± 0.05	0.01 ± 0.05	1.55 ± 0.03
4	1.44 ± 0.03	1.62 ± 0.02	0.10 ± 0.03	1.31 ± 0.03
6.25	1.50 ± 0.04	1.52 ± 0.05	0.03 ± 0.01	1.43 ± 0.04
8	1.48 ± 0.06	1.47 ± 0.04	0.01 ± 0.01	1.39 ± 0.04

* Values corresponding to dieldrin adsorption-desorption at 5°C (1) and 45°C (2).

• Values corresponding to dieldrin adsorption-desorption at salinity values of 25.56‰ (1) and 15.34‰ (2).

Table 2 Lindane adsorption-desorption partition coefficients and the resistant-reversible component partition coefficients to chitin in seawater (22°C and 36.52‰ salinity)

[Chitin] (g l ⁻¹)	K _a (l g ⁻¹)	K _d (l g ⁻¹)	K _o (l g ⁻¹)	K _x (l g ⁻¹)
2.5	0.080 ± 0.004	0.430 ± 0.002	0.039 ± 0.002	0.043 ± 0.002
4	0.067 ± 0.004	0.234 ± 0.028	0.028 ± 0.002	0.041 ± 0.003
6.25	0.062 ± 0.003	0.106 ± 0.004	0.018 ± 0.002	0.047 ± 0.001
6.25* ¹	0.079 ± 0.007	0.186 ± 0.003	0.032 ± 0.008	0.046 ± 0.001
6.25* ²	0.042 ± 0.002	0.043 ± 0.007	0.000 ± 0.001	0.044 ± 0.001
6.25* ¹	0.053 ± 0.003	0.075 ± 0.009	0.006 ± 0.002	0.049 ± 0.001
6.25* ²	0.049 ± 0.002	0.052 ± 0.004	0.001 ± 0.001	0.050 ± 0.001
7.5	0.060 ± 0.002	0.089 ± 0.006	0.012 ± 0.001	0.048 ± 0.002
10	0.055 ± 0.002	0.074 ± 0.004	0.009 ± 0.001	0.045 ± 0.002
12.5	0.048 ± 0.002	0.060 ± 0.002	0.009 ± 0.001	0.041 ± 0.004

* Values corresponding to lindane adsorption-desorption at 5°C (1) and 45°C (2).
* Values corresponding to lindane adsorption-desorption at salinity values of 25.56‰ (1) and 15.34‰ (2).

coefficients. This behaviour is affected by the chitin concentration and it is greater at lower chitin concentrations. The second effect is only observed for lindane. The lindane adsorption is affected by the adsorbent mass.

When after an initial desorption step we perform subsequent multiple desorptions the consecutive desorption isotherms are generated. Figure 4 shows the consecutive desorption isotherms for four dieldrin concentrations ($m = 1\text{ g l}^{-1}$ chitin) showing significant nonreversibility as it has been reported for other systems^{2,8,14}.

Gschwend and Wu¹⁵ and Baker *et al*¹⁶ have suggested that these complex sorptive phenomena are the result of artifacts associated with noncentrifugable or nonsettling

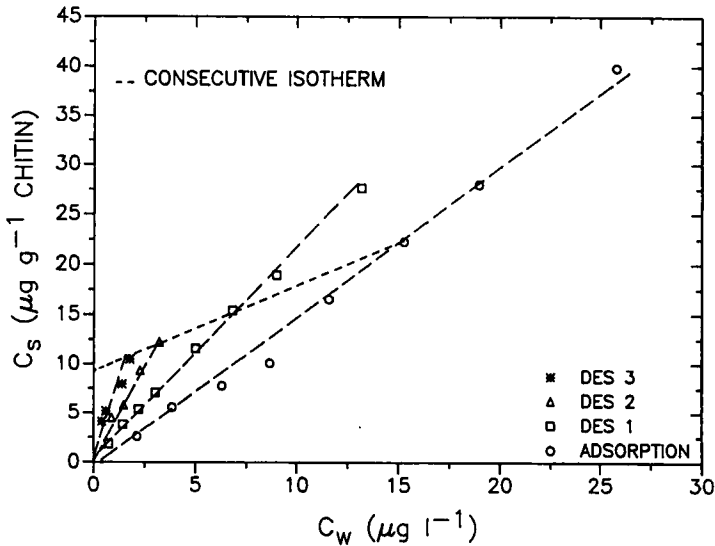


Figure 4 Single adsorption-desorption isotherms and consecutive isotherm of dieldrin on 4 g l⁻¹ chitin. From this Figure, the resistant-reversible componets of adsorption and desorption with the assumption of a linear consecutive desorption isotherm have been computed.

microparticles and organic macromolecules released from the solids. When experiments were carried out using water-washed chitin following Gschwend and Wu¹⁵ (approximately five washes), these do not show significant effects on the nonreversibility of the adsorption-desorption process. As in other research works^{17,18} we can conclude that these effects are not exclusively responsible for the nonsingular isotherms shown by lindane and dieldrin adsorbed to chitin. However, when other factors, such as temperature and salinity, were changed, the adsorption-desorption hysteresis observed was dramatically reduced as we show below.

Adsorption of dieldrin and lindane is expected to take place via various mechanism due to the very heterogeneous surface of chitin that accounts for the nonreversibility observed. The chitin surface sites may be divided, as a first approximation, in easily accessible surface sites and less accessible internal voids, which can attach pesticide with differing strength. Next, we attempt to describe the adsorption-desorption process using a reversible-resistant two-component model² which interprets incompletely reversible linear behaviour. Sorbed chemical, either at adsorption and at desorption equilibrium concentration is assumed to be the sum of a reversible sorbed component which readily adsorbs and desorbs reversibly and a resistant component which resists desorption over the time periods characteristic of experimental desorption equilibrium. The resistant component concentration C_o is defined as the extrapolated intersection of the consecutive isotherm and the ordinate, whilst the reversible component concentrations C_{xa} and C_{xd} corresponding to aqueous concentration C_{wa} and C_{wd} , respectively are defined as the difference between the adsorbed pesticide concentration at the adsorption and desorption points and the resistant concentration

$$C_{xa} = C_{sa} - C_o \quad (3)$$

$$C_{xd} = C_{sd} - C_o \quad (4)$$

The model assumes the reversible component concentrations C_x are described by a unique linear isotherm $C_x = K_x C_w$ both for the adsorption and desorption reversible components whilst the resistant component concentration follows a linear isotherm $C_o = K_o C_{wa}$. These isotherms define the reversible component partition coefficient K_x and the resistant component partition coefficient K_o .

Figure 5 shows for lindane adsorbed to 4 g l^{-1} chitin concentration the determined linear isotherm for the resistant and reversible component concentrations C_o , C_{xa} and C_{xd} , confirming the component model assumptions. Tables 1 and 2 show the results obtained for dieldrin and lindane, respectively, at all the chitin concentrations studied. These component partition coefficients K_x and K_o can also be calculated from the adsorption partition coefficient, K_a , and single desorption partition coefficient, K_d , by using the relationship²

$$K_o = \frac{mK_a(K_d - K_a)}{1 + m(K_d - K_a)} \quad (5)$$

$$K_x = \frac{K_a}{1 + m(K_d - K_a)} \quad (6)$$

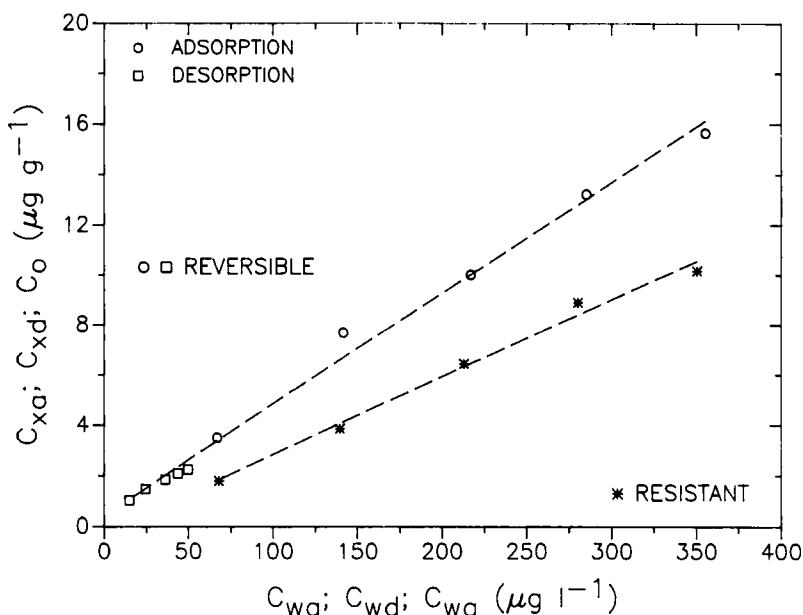


Figure 5 Resistant and reversible components and linear isotherms for lindane ($m = 4\ g\ l^{-1}$).

These theoretical values are completely in agreement with those obtained by extrapolating the experimental results which confirm the model used.

The effect of the chitin concentration on the partition coefficients has been explained by applying this model both to lindane and dieldrin adsorption-desorption processes.

When the lindane reversible and resistant component partition coefficients are plotted versus mass (Figure 6), it can be clearly observed that while the reversible partition coefficient is not dependent on the chitin concentration, $K_x = 0.045 \pm 0.004$, the resistant component partition coefficient is inversely related to chitin concentration, $K_o = v_o m^{-1}$, where $v_o = 0.0978 \pm 0.007$ is defined as a dimensionless distribution coefficient for the resistant component. Using these two known values, the dependence of the adsorption and desorption partition coefficients for lindane as a function of the adsorbent concentration yields

$$K_a = K_x + v_o m^{-1} \quad (7)$$

$$K_d = K_x + v_o m^{-1} [1 + (mK_x)^{-1}] \quad (8)$$

If we define the index of irreversibility as the difference of K_d and K_a ,

$$K_d - K_a = \frac{v_o}{m^2 K_x} \quad (9)$$

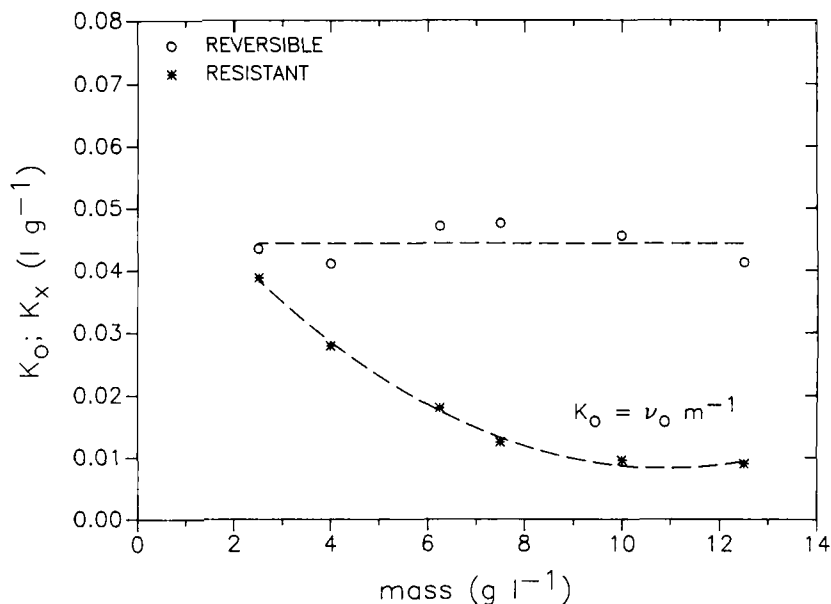


Figure 6 Lindane resistant, K_o , and reversible, K_x , component partition coefficient variation with adsorbent concentration. K_x is quite constant in the concentration range studied.

where

$$\begin{aligned} K_d - K_a &\rightarrow 0 & \text{as } m &\rightarrow \infty \\ K_d - K_a &\rightarrow \infty & \text{as } m &\rightarrow 0 \end{aligned} \quad (10)$$

These results explain the observed decrease of nonreversibility as the adsorbed mass concentration increase. It also gives the quantitative explanation for the almost reversible behaviour of the chitin at high concentrations.

When this same treatment is applied to dieldrin it is observed from Figure 7 that both reversible and resistant component partition coefficients are inversely related with the concentration

$$K_o = v m^{-1} \quad (11)$$

$$K_x = K_x^\infty - v m^{-1} \quad (12)$$

where $v = 0.3895 \pm 0.048$ is a dimensionless distribution coefficient for the resistant component and $K_x^\infty = 1.437 \pm 0.029 \text{ l g}^{-1}$, the partition coefficient describing the reversible adsorption-desorption of dieldrin to chitin.

Using these two parameters in the same way as before, we get

$$K_a = K_o + K_x = K_x^\infty \quad (13)$$

$$K_d = K_x^\infty + \frac{v}{m(mK_x^\infty - v)} \quad (14)$$

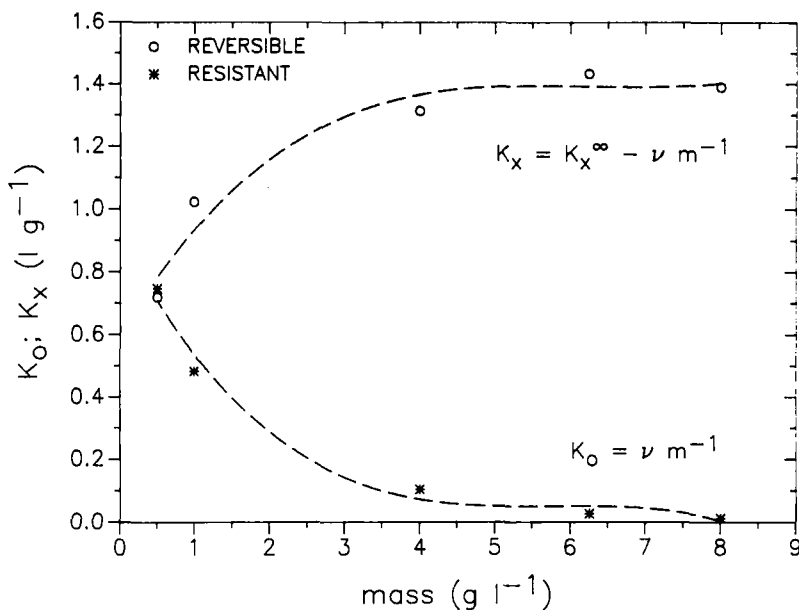


Figure 7 Dieldrin resistant and reversible component partition coefficient variation with chitin concentration. Both coefficients are inversely related with m (g/l).

It demonstrates, first, that the adsorption partition coefficient is independent of the mass of chitin as can be seen in Figure 2, and second, that for the desorption partition coefficient K_d , the second term of the sum accounts for the nonreversibility observed. At high chitin concentrations ($m > 6.25 \text{ g l}^{-1}$), both, the adsorption and desorption partition coefficients are the same as, and equal to K_x^∞ .

Effects of temperature, salinity and pH

According to the predicted value for an exothermic adsorption process, a decrease in the solution temperature, increases the adsorption process, both for dieldrin and lindane (Tables 1 and 2). Then again, an increase in temperature makes the process become reversible ($K_d = K_a$). This demonstrates that the nonreversibility observed for the adsorption-desorption of these pesticides on chitin in seawater is not due to experimental artifacts. If this is true, at high temperatures, we can expect more nonsettling microparticles and dissolved macromolecules that account for the nonreversibility shown. Thus, we can conclude that the nonreversibility behaviour is a result of the adsorption-desorption reaction mechanism.

When the two-component model is applied to both systems (Tables 1 and 2) we observe that only the resistant component is affected. An increase in the temperature decreases the resistant component towards zero. At high temperatures, the attractive forces associated with the resistant places between the pesticide and chitin may decrease⁹, resulting in greater desorption.

When the studies were carried out at different salinities (Tables 1 and 2), the adsorption of dieldrin is not affected by changes in the ionic strength of the aqueous solution. However, it is observed that a decrease in salinity makes the process become reversible ($K_a = K_d$ for $S = 15\text{‰}$). For lindane, both adsorption and desorption partition coefficients decrease as salinity decreases and at low salinities the process becomes reversible.

The application of the model to the experimental results shows for dieldrin (Table 1) that both, the resistant and the reversible component partition coefficients are affected by the ionic strength. The resistant component becomes zero while the reversible component partition coefficient reaches K_x^∞ as salinity approaches 15‰ . At lower salinity the adsorption-desorption reaction is not affected by changes in salinity. When the model is applied to the salinity effect on lindane adsorption-desorption (Table 2) it is observed that only the resistant component is affected, decreasing as salinity decreases.

Salinity may increase the adsorption as a result of the neutralization of the surface charge of chitin resulting from cation adsorption from seawater². This interpretation of the salinity data would appear to be consistent with the pH results. The adsorption reaction, both for lindane and dieldrin, was affected by changes in the pH solution values. An increase in the adsorption partition coefficients at pH values lower than 4.5 was observed due to changes in the charge characteristics of the chitin surface which should have favored maximization of positive surface charge. It can account for higher attractive coulombic interaction which can be responsible for the non-reversibility observed at the salinity values of seawater. At high salinity values the solid aggregation and particle size may also be affected and may increase the partitioning on the resistant sites and, therefore increase the resistant component partition coefficient.

CONCLUSION

It has been demonstrated that both dieldrin and lindane are adsorbed on chitin in seawater. The increased uptake of dieldrin over lindane may be due to the asymmetry of the dieldrin molecule which may increase the polarity compared to lindane. The increased polarity may result in stronger binding forces between the dieldrin and the chitin particles. Both pesticides show a nonreversibility behaviour at low chitin concentration, and for lindane the adsorption is affected by the chitin concentration. Both effects are not consequences of the presence of nonsettling chitin microparticles and dissolved organic macromolecules. This effect dramatically is reduced both by increasing temperature or by decreasing salinity, which may affect the adsorption-desorption mechanism. These types of behaviour have been explained taking into consideration that the adsorption is the result of two components, one resistant and another reversible. These two components can relate the variations of the adsorption and desorption partition coefficients with the adsorbent concentration by defining mass independent distribution coefficients. Thus, the reversible-resistant model explains most of the observed effects on the adsorption-desorption processes. However,

the mechanism that accounts for this behaviour is not clear yet, but the method described herein provides additional insight into the influence of aqueous-phase modifications (e.g., alteration of the pH, temperature and salinity).

References

1. R. H. Pierce, Jr., C. E. Olney and G. T. Felbeck Jr., *Geochim. Cosmochim. Acta* **38**, 1061–1073 (1974).
2. D. M. Di Toro and L. M. Horzempa, *Environ. Sci. Technol.* **16**, 594–602 (1982).
3. J. T. Coates and A. W. Elzerman, *J. Contam. Hydrol.* **1**, 191–210 (1986).
4. R. Haque, In *Dynamics, Exposure and Hazard Assessment of Toxic Chemicals*, R. Haque Ed. (Ann Arbor Science: Ann Arbor, MI, 1980).
5. L. L. Marking and R. A. Kimberle, *Aquatic Toxicology Proc. Second Annual Symposium on Aquatic Toxicology* (ASTMSTP 634; ASTM: Philadelphia, PA, 1977).
6. K. A. Lord, *Biochem. J.* **43**, 72–79 (1948).
7. P. Davar and J. P. Wightman, In *Adsorption from Aqueous solutions*, (P. H. Tewari, Ed. Plenum, New York, 1981) pp. 163–177.
8. D. E. Peck, D. L. Corwin and W. J. Farmer, *J. Environ. Qual.* **16**, 735–742 (1982).
9. D. M. Di Toro, L. M. Horzempa, M. M. Casey and W. Richardson, *J. Great Lakes Res.* **8**, 336–349 (1982).
10. T. C. Voice, C. P. Rice and W. J. Weber Jr., *Environ. Sci. Technol.* **17**, 513–518 (1983).
11. M. González-Dávila and F. J. Millero, *Geochim. Cosmochim. Acta* **54**, 761–768 (1990).
12. A. S. Y. Chau and B. K. Afghan, In *Analysis of Pesticides in Water* (CRC Press, Boca Raton, FL, 1982), Vol. II, Chap. 1, p. 11.
13. W. C. Stenn, D. F. Paris and M. Baughman, *Water Res.* **12**, 655–657 (1978).
14. P. A. Wahid and N. Sethunathan, *J. Agric. Food Chem.* **27**, 1050–1053 (1979).
15. P. M. Gschwend and S.-C. Wu, *Environ. Sci. Technol.* **19**, 90–96 (1985).
16. J. E. Baker, P. D. Capel and S. J. Eisenreich, *Environ. Sci. Technol.* **20**, 1136–1143 (1986).
17. P. R. Jaffé, *Environ. Sci. Health A21*, 55–69 (1986).
18. P. J. Witkowski, P. R. Jaffé and R. A. Ferrara, *J. Contam. Hydrol.* **2**, 249–269 (1988).